

**SEQUENCE SIMILARITIES BETWEEN TRYPTOPHAN SYNTHASE β SUBUNIT AND
OTHER PYRIDOXAL-PHOSPHATE-DEPENDENT ENZYMES**

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Summary: On the basis of 8 tryptophan synthase β subunits (EC 4.2.1.20) consensus patterns were constructed comprising two conserved motifs. Screening of the SWISSPROT protein sequence database with these patterns indicates similarities with O-acetylserine sulfhydrolases (EC 4.2.99.8), threonine synthases (EC 4.2.99.2), L- and D-serine dehydratases (EC 4.2.1.13/ EC 4.2.1.14) and threonine dehydratases (EC 4.2.1.16). Using multiple alignment procedures the similar regions could be extended. In connection with their pyridoxal-phosphate-binding-capacity and their positions in biochemical pathways evolutionary relationships among these enzymes are discussed. © 1990 Academic Press, Inc.

With the progress in sequencing techniques more and more primary structures of pyridoxal-phosphate-dependent enzymes become available. This offers the chance to get information about tertiary structures, functional mechanisms and evolutionary relationships. Since pyridoxal-phosphate is a cofactor of different enzymes like transaminases, decarboxylases, phosphorylases, dehydratases, synthases, sulfhydrolases etc. and since pyridoxal-phosphate is able to catalyse many of the respective reactions alone (without enzymes) the structures and sequences of these proteins can be expected to vary to a great extent and there is no reason for a common ancestor. This is confirmed by high resolution X-ray crystallography of glycogen phosphorylase A and B (1,2) some aspartate aminotransferases (mitochondrial isoenzyme from chicken heart (3), cytosolic isoenzymes from pig (4) and chicken heart (5)) and tryptophan synthase (6) for which no structural homology could be obtained. Other than, for instance, in nucleotide binding proteins (7) the binding sites (around the essential lysine) are different in the published tertiary structures (1,5,6).

Pyridoxal-phosphate-dependent enzymes are involved in many biochemical pathways, but an accumulation is found in anabolic reactions of the aspartate family and in connected pathways. Some homologies among pyridoxal-phosphate-dependent enzymes of the biosynthetic pathways of the aspartate family could be identified (8). Despite different functions they seem to have similar structures. Recently, distant homologies were found also between some aminotransferases (9), but the enzymes of this family act in a completely different way.

As a further application of our simple property pattern search method (10), which was also sensitive in identifying different nucleotide binding sites (11), we propose here a distant homology between tryptophan synthase β subunit and other pyridoxal-phosphate-dependent enzymes. Homologies among this family of pyridoxal-phosphate-dependent enzymes allow structural and functional insights in the mechanisms of all these enzymes and provide some hints related to the evolution of biochemical pathways.

Materials and Methods

By means of FASTP (12) the sequences most similar to tryptophan synthase β subunit of salmonella typhimurium (with known structure) could be identified in SWISSPROT protein sequence database (release 12.0, containing 12305 sequences). Significant matches were given by tryptophan synthase β subunits from other species and the equivalent part of the uncleaved yeast enzyme. The 8 sequences found in this way were aligned using our program MULTIS (13) which allows to compare up to 15 protein sequences now.

In tryptophan synthase pyridoxal-phosphate is covalently bound to an essential lysine of the β subunit (or to the equivalent region of the uncleaved enzymes). A second conserved region is a glycine rich turn which ligates through H-bonds the phosphate group of pyridoxal-phosphate (6). For these regions property patterns were constructed using PAT (14). This procedure leads to rows of amino acids in each position of a sequence segment (fig.1b). With these patterns the SWISSPROT database was screened.

Since our simple property pattern procedure is able to recognize also distantly related proteins (10), for multiple alignment of protein sequences with a low degree of amino acid identity we have developed an additional program PULIN, which carries out the multiple alignment via an iterated profile alignment (15-17) using the Dayhoff PAM250 matrix (18). We used PULIN to align those proteins, for which some similar regions were detected by our property pattern method.

Results

When the SWISSPROT was screened with our property pattern comprising the two motifs described above in addition to the

a)

| codes | proteins | lysine-motif | | glycine rich turn | |
|-------------|-------------------------------------|--------------|------------------------------|-------------------|----------------------------|
| | | pos m | sequence segments | pos m | sequence segment |
| CYSK\$ECOLI | O-acetylserine sulfhydrolase | 25 | 3 NGRILAKVESRNP-SFSvKCRiGAN | 167 | 3 GQVDVfIAGvGTGGTLTGvRYI |
| CYSK\$SALTY | : | 25 | 3 NGRILAKVESRNP-SFSvKCRiGAN | 167 | 3 GQVDVfIsgVGTGGTLTGvTRYI |
| SDHD\$ECOLI | D-serine dehydratase | 100 | 2 SGQLLLKKDShLPISSiKaRGGIy | 269 | 1 DNPLFVYLPCGvGGGPGGvAFGL |
| SDHL\$RAT | L-serine dehydratase | 24 | 1 GTSVFLKMDSSQP-SGSFKIRGiH | 194 | 0 AKPGAIVLsvGGGGLLcGVvQGL |
| THD1\$ECOLI | threonine dehydratase | 45 | 0 DNVILVKREDRQP-VHSFKLRGAYa | 178 | 1 AHLDRVfVPVGGGLAAGvAVI |
| THD2\$ECOLI | : | 41 | 0 KGEIFLKFENMQR-TGSFKIRGAfN | 174 | 0 YDvDvNVIvPIGGGGLIAGIvAVI |
| THDH\$YEAST | : | 92 | 0 NTNvILKREDLLP-VFSFKLRGAYn | 228 | 0 NKIGAVfVPVGGGGLIAGIvAYL |
| THRC\$BACSU | threonine synthase | 42 | 1 GIELHVkTEGvNP-TGSFKDRGMvM | 175 | 0 EAPDvLAIvPVGNAGNITAYvKGF |
| THRC\$BRELA | : | 42 | 0 GVQLYGKYEgANP-TGSFKDRGMvM | 175 | 0 NAPDvLAIvPVGNAGNITAYvKGF |
| THRC\$ECOLI | : | 90 | 2 ESDvGCIEIvFHGP-TLAFKDFGGfR | 238 | 3 RNQLvVSVpSGNfGDLTAGLLAK |
| TRP\$YEAST | tryptophan synthase | 367 | 0 GAQIvLKREDLNH-TGSHKINNALa | 520 | 1 KLPDAvVACvGGGSNSTGMfSFF |
| TRPB\$BACSU | tryptophan synthase β subunit | 73 | 0 GAKIvYLKREDLNH-TGSHKINNALg | 225 | 0 TMPDKvVACvGGGSNAMGMfGAF |
| TRPB\$BRELA | : | 82 | 1 fARIvFLKREDLVH-GGAHKTNQvIG | 234 | 1 KLPDvVVACvGGGSNAIGMfADF |
| TRPB\$CAUCR | : | 82 | 0 GAKIvYfKRDELNH-TGSHKINNALg | 234 | 0 RLPDAvVACIGGGsNAIGLFHfPF |
| TRPB\$ECOLI | : | 69 | 0 NTTLYLKREDLLH-GGAHKTNQvLG | 221 | 0 RLPDAvIACvGGGSNAIGMfADF |
| TRPB\$PSEAE | : | 76 | 0 GAKIvYLKREELNH-TGAHKINNCIG | 228 | 0 RLPDSvLVACIGGGsNAMGLFhFP |
| TRPB\$PSEPU | : | 78 | 0 GAKIvFFKREELNH-TGAHKVNNCIG | 230 | 0 RLPDSvLVACvGGGSNAMGLfHEF |
| TRPB\$SALTY | : | 69 | 0 RTTLYLKREDLLH-GGAHKTNQvLG | 221 | 0 RLPDAvIACvGGGSNAIGMfADF |

b) corresponding allowed amino acids:

| | |
|---------------------------|-------------------------|
| AAAIAAKEDAAACIAAAFKCCAAAA | AACAACAACGAAAAAAAAAAAF |
| CCCLCC FECCCF-CCSH DGCCCC | CCICCFCCI CGGCCGCCCCI |
| DGDVFF H DFGH GF I EHGGFF | DDLDFIFGNL G SDIG FFDDL |
| EIE GG K EHIK NG K IKMIGG | EEMEHLIIPM N ELI GGEEM |
| GLI HI M FILM SH L LMNLHM | FFNGIMLLSV S GML IIFF |
| KMK IL Q GKMN TI M MNGMIN | GGPIKvMMT T INM LLHG |
| MNL KM R MLNP VK W NQSVKQ | HHQLL NV LPT MMIM |
| NQM LV W NMQR L Y QRT LS | KITMM Q MQV TTKN |
| QSN M Y QNSR M TT MT | MKVNN S NS VVLP |
| RTQ T SQTT T VW NW | NL GQ T QT WWMQ |
| SVR V TRVW V Y QY | QM SR V SV YYQS |
| T S W WS Y W R | RN TS W T RT |
| T Y YT Y S | SQ VT Y V SW |
| V V T | TR V Y TY |
| W V | WS W V |
| Y W | YT Y W |
| | V Y |

Fig. 1. Lysine-motif and glycine rich turn of pyridoxal-phosphate-dependent enzymes.

tryptophan synthases also the proposed pyridoxal-phosphate binding sites (7) of 2 threonine dehydratases, 2 threonine synthases and a serine dehydratase match the pattern with only a few deviations. The respective regions of the detected enzymes were used to improve our property pattern and the SWISSPROT was screened again. With this second run all threonine synthases, threonine dehydratases and serine dehydratases of the SWISSPROT were found. In addition 2 O-acetylserine sulfhydrolases were detected for which similarities of their C-terminal part to tryptophan synthase β subunit were described (19). Fig.1a shows the results of this run. Up to 5 mismatches (deviations) from the first and up to 4 from the second motif were allowed, but out of 12305 proteins in SWISSPROT only the proteins shown in fig.1a match both motifs. With a third run on the bases of property patterns derived from all regions shown in fig.1, no further sequence segment was detected before "noise" appeared.

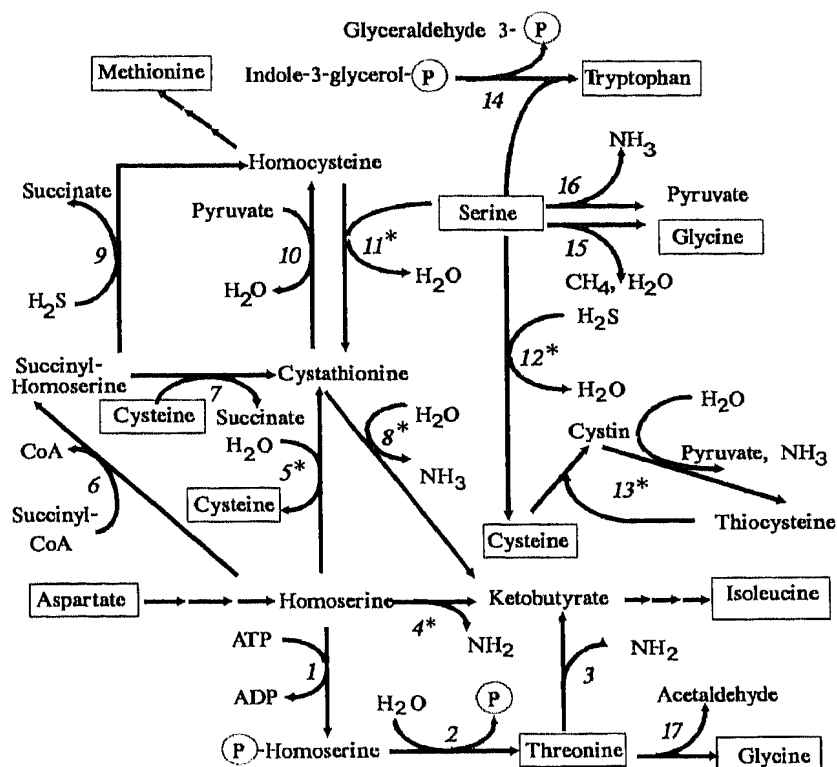


Fig. 3. Metabolic map of amino acids and pyridoxal-phosphate-dependent enzymes.

To verify the indicated similarities among the detected enzymes we have used our multiple alignment program PULIN. The similar regions could be extended and we propose now an alignment, which reveals a similar folding topology in long segments of the included enzymes (fig.2).

The similar sequence segments agree with structural findings, because deletions and insertions mostly occur in regions between corresponding secondary structure elements of tryptophan synthase (fig.2). Our alignment with O-acetylserine sulfhydrylase is somewhat different to that proposed by Levy and Danchin (19). So we have related the potential pyridoxal-phosphate-binding site in O-acetylserine sulfhydrylase to a region in tryptophan synthase around the lysine-87 (numbering according to the enzyme from salmonella typhimurium, fig.2) as indicated by our property patterns (fig.1).

Only in 3 positions amino acids were found to be absolutely conserved in all sequences (marked by an exclamation mark in fig.2). One of them is the essential lysine covalently bound to pyridoxal-phosphate, the second is a glycine of the second motif used for the pattern construction and the third is located in a

Fig. 2. Conserved sequences among pyridoxal-phosphate-dependent enzymes.

region surprisingly found to be the most conserved one corresponding to residues 112 to 134 in tryptophan synthase from salmonella thyphimurium (fig.2). This region comprises an α -helix with a subsequent turn and consists of a cluster of tiny amino acids. Further conserved regions comprise the motifs used for pattern construction. All three most conserved regions are not located in the β -sheet core so that they seem to have essential functions in pyridoxal-phosphate-binding.

Discussion

The pairwise identities between enzymes of the different subgroups in fig.2 range between 15% and 19%. These values are typical for the so called 'twilight zone' in which usually automatic alignment procedures fail (20,21), but background information argues in favour of their similarities. All enzymes involved in our alignment and proposed to be homologous have some features in common: (i) they all need pyridoxal-phosphate as a cofactor, (ii) there are some similarities in their reaction mechanisms via an intermediate and (iii) they all catalyse successive steps in biochemical pathways (fig.3). The third point led to the hypothesis of an evolution of these pathways from an ancestor pyridoxal-phosphate-binding protein. In fig.3 some successive reactions of the aspartate family and their connections to tryptophan synthase are shown. All of them are catalysed by pyridoxal-phosphate-dependent enzymes. Therefore we have also investigated other enzymes of this scheme (fig.3) not found by our pattern and alignment procedures. Homologies between cystathionine γ -synthase (EC 4.2.99.9) and cystathionine β -lyase (4.4.1.8) were detected (22) and we have also found a high level of similarity to O-acetylhomoserine thiolase (EC 4.2.99.10). O-acetylhomoserine thiolase can catalyse the same reaction as O-acetylserine sulfhydrylase which is, in turn, homologous to tryptophan synthase. Despite an amino acid identity of 11% (10 gaps were introduced by PULIN, data not shown) a homology could not be verified. The identical positions are in contrast to the alignment of O-acetylhomoserine thiolase with cystathionine β -lyase and cystathionine γ -synthase.

Nevertheless, the evolution of these pathways can be explained as a series of gene duplication events. In various aminotransferases or decarboxylases only the region responsible for the recognition of the respective amino acid had to be changed

after gene duplication to explain the different positions in the pathways. In contrast, the family of similar pyridoxal-phosphate-dependent enzymes discussed here reveals very early gene duplication events leading to the development of biochemical pathways.

The suggested homology between tryptophan synthase (β subunit) and the other enzymes presented here can be used to build topology models of all these proteins. The conserved regions and positions show that despite the expected variability of pyridoxal-phosphate-dependent enzymes (see introduction) there seems to exist only a limited set of protein topologies compatible with pyridoxal-phosphate-binding via lysine.

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