

Domains in plexins: links to integrins and transcription factors

Integrins are adhesion molecules that bind diverse cell-surface and extracellular-matrix ligands¹. They are heterodimeric receptors, containing α and β subunits. No significant sequence similarity between the extracellular domains of integrin β subunits and any other protein has been reported, although the presence of a DXSXS motif, and secondary-structure predictions, suggests that the most-conserved region adopts an I-domain-like fold (also called a von Willebrand factor A domain)²⁻⁴. Furthermore, the C-terminal third of the extracellular domain contains four internal, cysteine-rich repeats⁵. The only other cysteine-rich region (a cluster of seven cysteine residues in 50 residues) is in the N-terminal segment of mature integrin β subunits. Six of these cysteines form disulfide bonds to one another; the remaining, first, cysteine forms a long-range disulfide bond that links the N-terminal and C-terminal cysteine-rich regions⁶.

PSI-BLAST searches⁷ with the N-terminal cysteine-rich region of individual integrin β subunits retrieve a region in a mouse neuronal cell-surface molecule, plexin 2 (Ref. 8), at the third iteration ($E = 3 \times 10^{-4}$). Further iterations reveal homology to previously described internal repeats of this region in plexins^{8,9}. Subsequent PSI-BLAST searches with this repeat reveal in the first iteration significant homology to a family of proteins that act as 'semaphores' for growth-cone guidance¹⁰, the semaphorins ($E = 3 \times 10^{-4}$). In later iterations, proteins related to a signaling receptor, mahogany, that functions in the brain as a suppressor of obesity are retrieved¹¹ (E values are of the order of 10^{-4}). Previously, it has been shown that the three repeats in plexin are homologous to a small region of the hepatocyte growth factor (HGF) receptor,

MET (Ref. 9), and also to the virus-encoded semaphorin receptor (VESPR)¹². Reciprocal studies using both sequence profiles¹³ of the repeats from the plexin family and PSI-BLAST searches with several members of the major subfamilies indicate that other proteins (such as MEGF8 or C21orf1) contain the repeat (Fig. 1). We named this region the PSI domain (after the better-characterized families plexins, semaphorins and integrins). The PSI domain is part of the original definition of the sema domain¹⁰, but further studies of the modular architecture of the semaphorins reveal that there are indeed semaphorins that lack the PSI domain, such as A39R from vaccinia virus. This leads to a redefinition of the original sema domain, and only this

redefined sema domain seems to be a marker for semaphorins.

The PSI domains of the proteins shown in Fig. 1 are ~50 residues in length and usually contain eight cysteine residues. On the basis of experimental determination of disulfide bonds in integrins⁶, we can predict the location of disulfide bonds (between cysteine two and cysteine four, between cysteine three and cysteine eight, and between cysteine five and cysteine seven) for the entire family (Fig. 2a). The remaining two cysteines (cysteine one and cysteine six), when both are present, are also predicted to be disulfide bonded; in integrins, cysteine one forms the long-range disulfide bond⁶, and cysteine six is absent. In several family members, cysteine five and

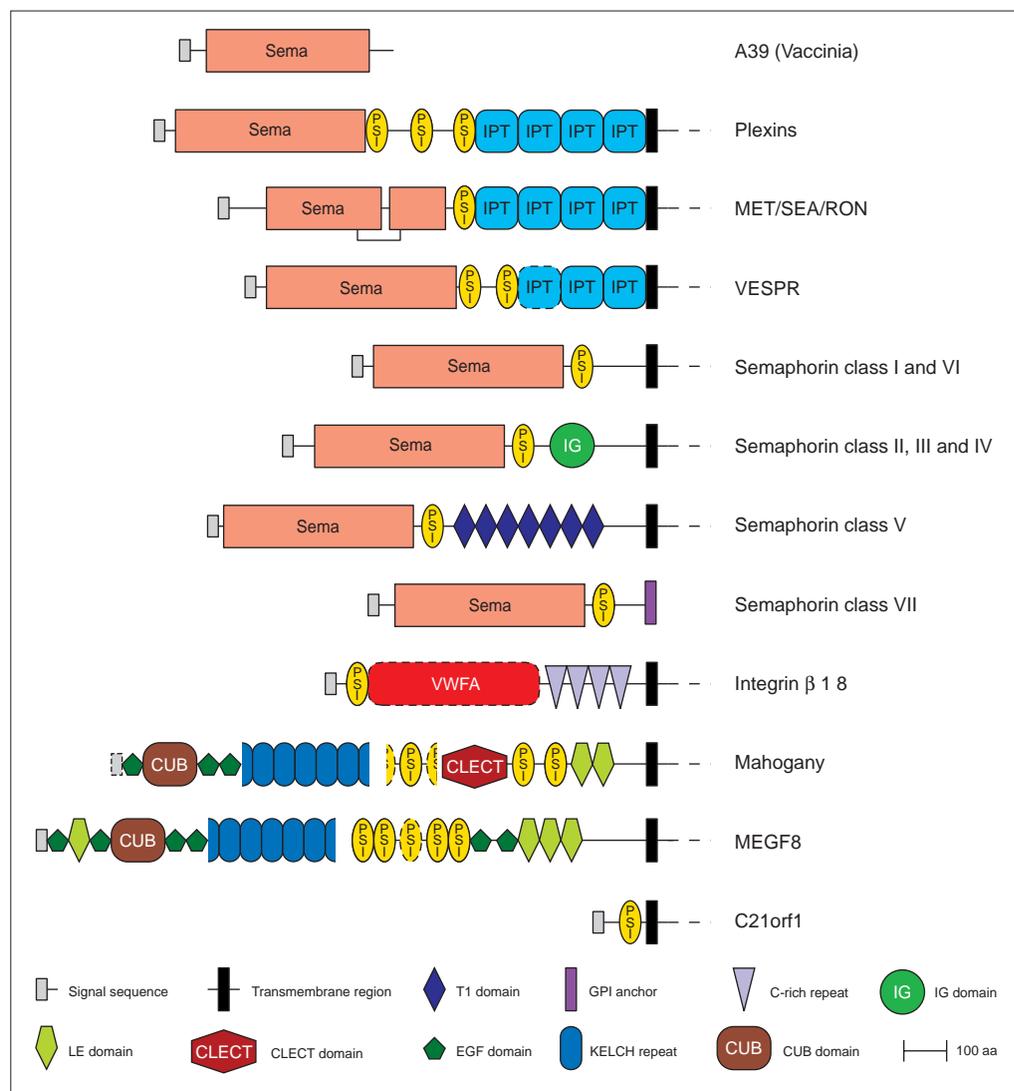


Figure 1

The extracellular region of proteins containing PSI and IPT domains. Only regions of proteins that have distinct modular organizations to their extracellular regions are shown. Domain names are according to Ref. 21. A broken line surrounding a domain indicates that it does not give significant E values, but its presence is supported by context information. The cysteine (C)-rich repeats in the integrins somewhat resemble epidermal growth factor (EGF)-like domains, although they contain two additional cysteines. aa, amino acid residues; GPI, glycosylphosphatidylinositol; IG, immunoglobulin; LE, laminin epidermal growth factor-like; VESPR, virus-encoded semaphorin receptor; VWFA, von Willebrand factor type A domain.

Figure 2

Multiple alignments of selected members of the PSI- and IPT-domain families. The names of the proteins (multiple domains in the same proteins are labeled a, b, c or d), the species and the number of the residue at the start of the domain are shown on the left. Database accession numbers are shown on the right. Conserved cysteine residues are shown in red; the conserved tryptophan residue is shown in blue; conserved hydrophobic residues are shown in green; other conserved residues are shown in bold. The consensus sequence shown below contains conserved features in the domain: C and W denote conserved cysteine and tryptophan residues; t and h indicate turn-like/polar and hydrophobic positions, respectively. Predicted secondary structure (sec. struct. pred.)¹⁴ is also shown (h, helix; E, β -sheet predicted with high significance; e, β -sheet predicted with lower significance). **(a)** Multiple alignment of different PSI domains. Cysteine residues are numbered above the alignment and color-coded on the basis of predicted disulfide-bonded residues⁶. The predicted secondary structure of the PSI domain is a consensus derived from independent results for plexins, semaphorins, integrins and mahogany-like domains. **(b)** Multiple alignment of different IPT domains. The known secondary structure of the D domain of *Bacillus stearothermophilus* (bs) cyclodextrin glucanotransferase (1CYG) is shown below the alignment. ce, *Caenorhabditis elegans*; gg, *Gallus gallus*; hs, *Homo sapiens*; mm, *Mus musculus*; tc, *Tribolium confusum*; xl, *Xenopus laevis*.

cysteine seven are both missing, which provides additional support for the idea that the two residues are normally bonded together (Fig. 2a). Secondary-structure predictions¹⁴ strongly indicate that an α -helix is present between cysteine seven and cysteine eight (Fig. 2a) and thus support the alignment shown in Fig. 2a.

The presence of multiple PSI domains, and of a (redefined) sema domain, classifies plexin, MET and VESPR (Fig. 1) as semaphorins. Moreover, it leaves only the region N-terminal to the transmembrane region in the extracellular part of plexins undescribed. PSI-BLAST searches with this region suggest that it has a repeat-like character. For example, if a search is initiated with the second repeat of RON_HUMAN (Fig. 2b), not only three internal repeats but also regions in MET and SEX are significantly similar (E values are 10^{-10} and 10^{-6} , respectively). Further iterations reveal homology to Olf1/Ebf-like transcription factors (e.g. for OLF-1, $E = 10^{-4}$) and to the nuclear factor of activated T cells (NFAT) family of transcription factors. Wisetool profiles¹³ and Hidden Markov Model¹⁵ searches based on the three repeats of the motif in plexins and MET detect a fourth repeat in these proteins, two repeats in VESPR and one in the hypothetical protein YMS5_CAEEL (Fig. 2b). A BLAST search with the motif of the transcription factor XCOE2, a close homolog of OLF-1, retrieves the immunoglobulin-like (D) domain of *Bacillus stearothermophilus* cyclodextrin glucanotransferase, whose structure is known (PDB accession number: 1CYG) with a significant E value ($E = 3 \times 10^{-4}$). We therefore propose the name IPT (for immunoglobulin-like fold shared by plexins and transcription factors) for this domain (Fig. 2b).

Within the semaphorin family, the IPT domain is present only in plexins, MET and VESPR, but the PSI domain is associated with many more family

members. Although most seem to have a role in neuronal development, several family members appear to have immunological functions¹⁶. Mahogany¹¹ is the only other PSI-domain-containing protein for which functional information is currently available. It is a signaling receptor and functions in the brain as a suppressor of obesity¹¹. A receptor-like architecture for most of the proteins that contain the PSI domain is thus the only clear common theme, although soluble forms exist – at least in the cases of some semaphorins¹⁶ and mahogany¹¹. A monoclonal antibody that binds to the PSI domain of the integrin $\beta 2$ subunit does not block ligand binding, but monoclonal antibodies to the adjacent, putative, I-like domain do (Ref. 17, and C. Huang and T. A. Springer, unpublished). MET (Ref. 18) and some semaphorins^{19,20} dimerize, and the PSI domain might be involved in this process. Plexins can also bind to plexin molecules on other cells and, thereby, mediate cell adhesion through a homophilic binding mechanism⁹.

In summary, currently, the PSI domain can be described, only vaguely, as an extracellular, putative, protein-binding domain, but its detection in this family of proteins should enable structural studies that provide further insights into the functions of the proteins shown in Fig. 1 (see also Box 1).

Box 1. Note added in proof

After submission of this manuscript, Winberg *et al.*²² reclassified semaphorins by including plexins, VESPR and MET, in which they also noted G-P motifs (part of the IPT domain). Furthermore, a similarity between some transcription factors and plexin has been noted recently²³.

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References

- Hynes, R. O. (1992) *Cell* 69, 11–25
- Lee, J.-O., Rieu, P., Arnaut, M. A. and Liddington, R. (1995) *Cell* 80, 631–638
- Tozer, E. C. *et al.* (1996) *J. Biol. Chem.* 271, 21978–21984
- Tuckwell, D. S. and Humphries, M. J. (1997) *FEBS Lett.* 400, 297–303
- Tamkun, J. W. *et al.* (1986) *Cell* 46, 271–282
- Calvete, J. J., Henschen, A. and González-Rodríguez, J. (1991) *Biochem. J.* 274, 63–71
- Altschul, S. F. *et al.* (1997) *Nucleic Acids Res.* 25, 3389–3402
- Kamayama, T. *et al.* (1996) *Biochem. Biophys. Res. Commun.* 226, 396–402
- Ohta, K. *et al.* (1995) *Neuron* 14, 1189–1199
- Kolodkin, A. L., Matthes, D. J. and Goodman, C. S. (1993) *Cell* 75, 1389–1399
- Nagle, D. L. *et al.* (1999) *Nature* 398, 148–152
- Comeau, M. R. *et al.* (1998) *Immunity* 8, 473–482
- Birney, E., Thompson, J. D. and Gibson, T. J. (1996) *Nucleic Acids Res.* 24, 2730–2739
- Rost, B., Sander, C. and Schneider, R. (1994) *CABIOS* 10, 53–60
- Eddy, S. R. (1998) *Bioinformatics* 14, 755–763
- Zhou, L. *et al.* (1997) *Mol. Cell. Neurosci.* 9, 26–41
- Huang, C., Lu, C. and Springer, T. A. (1997) *Proc. Natl. Acad. Sci. U. S. A.* 94, 3156–3161
- Gaudino, G. A. *et al.* (1994) *EMBO J.* 13, 3524–3532
- Koppel, A. M. and Raper, J. A. (1998) *J. Biol. Chem.* 273, 15708–15713
- Klostermann, A. *et al.* (1998) *J. Biol. Chem.* 273, 7326–7331
- Bork, P. and Bairoch, A. (1995) *Trends Biochem. Sci.* 20, poster C02
- Winberg, M. L. *et al.* (1998) *Cell* 95, 903–916
- Aravind, L., Dixit, V. M. and Koonin, E. V. (1999) *Trends Biochem. Sci.* 24, 44–53